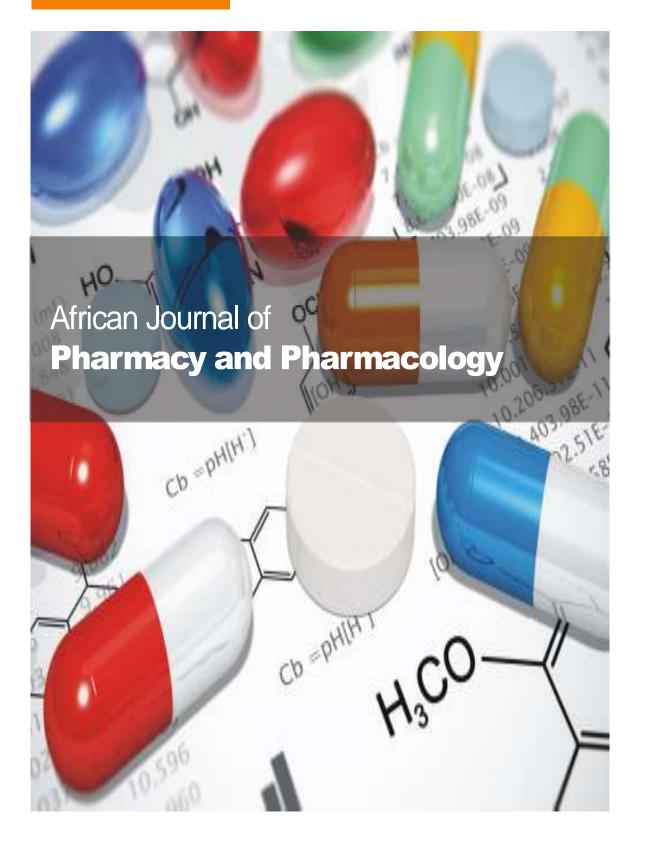
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African Journal of Pharmacy and Pharmacology

Full Length Research Paper

Physician prescription practice of antibiotics for upper respiratory tract infection at Kilimanjaro Christian Medical Centre Moshi, Tanzania

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Upper respiratory tract infection occurs commonly in both children and adults and is a major cause of morbidity worldwide. Inappropriate antibiotic prescription for upper respiratory tract infections is associated with increasing antibiotic resistance, healthcare costs, adverse events, and poor patient outcomes. The objective of this study was to determine physician prescription practices of antibiotics for upper respiratory tract infections at Kilimanjaro Christian Medical Center hospital in Moshi, Tanzania. This was a retrospective hospital-based cross sectional study which systematically sampled files of patients with diagnosis of upper respiratory tract infection. Information from a total of 300 patients' prescriptions were collected, reviewed and analyzed. The most common infections diagnosis was non-specific upper respiratory tract infections accounting for 102 (34.0%) followed by rhinitis and tonsillitis both accounting for 52 (17.3%) with the least being common cold 22 (7.3%). Antibiotics were prescribed to 200 (66.7%) patients with upper respiratory tract infections. Amoxicillin alone was the most preferred drug for all upper respiratory tract infections 91 (31.5%). In the multivariable logistic regression analysis, patients with cough and running nose (AOR=16.41, 95% CI: 1.95-138.19) had higher odds of being prescribed with antibiotic as compared to those without such symptoms (AOR=1.98, 95% CI: 1.04-3.77), respectively. Antibiotics are being over-prescribed among patients with upper respiratory tract infection. Interventions to reduce the over-prescription and hence overuse of antibiotics for upper respiratory tract infections are urgently needed.

Key words: Antibiotics prescribing, upper respiratory tract infection, Tanzania.

INTRODUCTION

Upper respiratory tract infections (URTIs) is a term used

to describe acute infections involving the nasal cavity,

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pharynx and larynx (Mungrue et al., 2009; Yoon et al., 2017). URTIs are classified according to the area of inflammation, which are tonsillitis, pharyngitis, laryngitis, sinusitis, common cold, influenza and otitis media (Centre for Clinical Practice, 2008). Moreover, Otitis media has not been classified as URTIs, although it has been linked with the upper respiratory tract, since it occur as a complication of URTIs hence tends to be identified within URTIs (Chonmaitree et al., 2008).

Most of the URTIs are of viral etiology and these include viruses in the family rhinovirus, coronavirus, parainfluenza, respiratory syncytial virus, adenovirus and influenza (Cotton et al., 2008). Occasionally, bacteria may develop after a viral illness such as a cold or the flu. The significance of pathogenic bacteria in the upper respiratory tract is yet to be characterized (MacIntyre et al., 2017), since positive nasopharyngeal bacterial culture is a weak predictor of URTIs because healthy individuals often carry pathogenic bacteria (Ouédraogo et al., 2014). People are generally thought to be asymptomatic carriers of bacteria (Faden et al., 1997; Givon-lavi et al., 2002; Bogaert et al., 2004). Most episodes of URTIs are typically self-limiting, thus do not require physician visit or antibiotic(s) prescription (Peroš-Golubičić and Tekavec-Trkanjec, 2015; Llor et al., 2017). Acute respiratory infections which are divided into URTI and lower respiratory tract infections, have been established as one of the leading causes of childhood morbidity and mortality in Africa (Symekher et al., 2009). It has been estimated that up to 1.9 million children die each year from acute respiratory with nearly 70% of deaths occurring in Africa and South East Asia (Simoes et al., 2006; Symekher et al., 2009). However, the burden of URTI in most African countries including Tanzania has not been documented.

Despite consistent and continued education among healthcare professionals on antibiotic resistance. antibiotic prescribing rates for URTIs remain high in general practice (Fletcher-Lartey et al., 2016). World Health Organization (WHO) estimated that up to 60% of people with URTIs receive antibiotics inappropriately (Kunda et al., 2015). Inappropriately use of antibiotics contributes to the emergency of antimicrobial resistance (AMR), which is a major public health problem worldwide. Documented factors associated with antibiotic prescription for URTI by physicians include fever (temperature >38°C), fear of complications, inflamed eardrums, cough, and throat irritation as well as primary caregivers' pressure and patients financial gains (Al-Enezi et al., 2011).

Guidelines by Centre of Disease Control (CDC) do not recommend antibiotic prescribing in non-specific URTIs because antibiotics neither enhance illness resolution nor prevent complications (CDC, 2017). One study showed that only 12% of physician would request laboratory tests such as culture and sensitivity before prescribing antibiotics, and 88% of the physician considered laboratory investigations as unnecessary (Mohan et al., 2004). Misdiagnosis and improper diagnosis leads to incorrect antibiotics prescription and so increases AMR in many parts of the world (Haque, 2017). For better treatment outcome, healthcare workers are encouraged to request for microbiological testing prior to start of antibiotics for a more rational antibiotic use in both children and adults (Levy-Hara et al., 2011; Chaw et al., 2018).

Several studies have shown the rate of antibiotic prescriptions in relation to the diagnosis of URTIs made which was high and inappropriate (Teng et al., 2004; Gwimile et al., 2012; Kunda et al., 2015). Inappropriate antibiotic prescription for URTI is a global public health problem, therefore URTIs are important for strategies aimed at reducing excess antibiotic use because antibiotics are frequently prescribed in these illnesses that are predominantly of viral etiology (Easton and Saxena, 2010; Kunda et al., 2015; Fletcher-Lartey et al., 2016; Zhang et al., 2017).

Although prescribing patterns generally differ between countries due to the national guidelines and drugs available, very little is known about antibiotics prescribing practices in Tanzania and specifically at Kilimanjaro Christian Medical Centre (KCMC). Despite the fact that awareness of the consequences of antibiotic misuse is increasing among population (Mbwambo et al., 2017), as well as among healthcare providers (Lyimo et al., 2018), in the northern part of Tanzania, overprescribing of antibiotics is still being practiced at high rate even before availability of laboratory results are made available (Chilongola et al., 2015; Kajeguka et al., 2017). Therefore, this study aimed at assessing physician prescription practice on antibiotics for URTIs.

METHODS

Study design and area

This was a retrospective hospital-based cross sectional study carried out from April 2017 to July 2017 at KCMC hospital in Moshi, Kilimanjaro region, Tanzania. Physicians' antibiotic prescriptions from January 2015 to June 2017 were included. KCMC is a consultant referral hospital with 630 inpatient beds serving several regions in northern part of Tanzania. The study was conducted in three departments; which are Pediatric, Internal medicine and Outpatient department (OPD).

Sample size determination and sampling

The following formula was used to obtain the required sample size:

$$N = [Z^2 P (1-P)] / (d^2),$$

where N is the required sample, Z is the confidence level at 95% (1.96), P is the prevalence of 0.78, and d is the margin of error at 5% (0.05).

Variable		Measure
Age in years, Mean ± SD		2.02 ± 1.03
	<15	126 (42.0)
	15-35	84 (28.0)
Age categories (in years)	36-45	25 (8.3)
	>45	65 (21.7)
0	Male	92 (30.7)
Sex	Female	208 (69.3)
	Pediatric	62 (20.7)
Ward/Department	Internal medicine	42 (14.0)
	Outpatient Department	196 (65.3)

Table 1. General characteristics of study participants (N=300).

The prevalence of antibiotics prescription on URTI was 78% (Kunda et al., 2015). A maximum of 300 patient files were systematically sampled. Records with missing data on URTIs were excluded.

Data collection methods and tools

All prescriptions of patient with the diagnosis of URTI from physician in selected departments/wards at KCMC hospital were included. Data was extracted from patient's medical records (files). The following data were recorded; age, sex, diagnosis given, if laboratory test were requested, if antibiotic was prescribed, type of antibiotic prescribed, number of antibiotics prescribed and type of URTIs. Signs and symptoms that prompted antibiotic prescription were recorded, such as fever (Temperature >38°C), chest pain, cough, running nose, exudates in throats, inflamed ear and difficulty in breathing.

Data analysis

Data were analyzed using IBM SPSS Statistics for Windows, Version 22.0 (IBM Corp, Armonk, NY, USA). Descriptive statistics was used to summarize data. Differences between categorical data were calculated using Pearson's Chi-square test (χ^2). Factors that were found to have a level of significance of p≤0.05 were then entered into the final model of the multivariable logistic regression analysis, which was used to compute adjusted odds ratio (AOR) and 95% confidence intervals (95% CI) to assess the independent associations of these variables with outcome of interest (antibiotic prescription). A p<0.05 was significant.

RESULTS

Demographics characteristics and URTIs diagnosed

A total of 300 patient files were reviewed. The mean age in years (\pm Standard deviation) was 2.02 \pm 1.03, with

those aged 15 years and below constituting the highest proportion 126 (42.0%) followed by those aged 16 to 35 years 84 (28.0%). Two hundred and eight participants (69.3%) were female. Most of the participants were from OPD 196 (65.3%), followed by pediatric 62 (20.7%) and internal medicine 42 (14.0%) (Table 1). The most common URTIs diagnosed were non-specific URTIs 102 (34.0%) followed by rhinitis and tonsillitis, both 52 (17.3%). The least diagnosis assigned was common cold 22 (7.3%) (Figure 1).

Antibiotics prescribing pattern for URTIs

Two hundred patients with URTI (66.7%) received atleast one antibiotic (Any antibiotic); with the highest in pharyngitis 47 (100%), followed by non-specific URTIs 82 (80.4%) and rhinitis 38 (73.1%) (Table 2). Amoxicillin alone was the most frequently prescribed antibiotic for common cold 16 (72.7%). Ampicillin was frequently prescribed for otitis media 7 (28.0%), followed by tonsillitis 14 (26.9), and pharyngitis 11 (23.4%). Ampiclox was commonly prescribed for otitis media 10 (40.0%) (Table 2).

Signs and symptoms that influenced antibiotic prescription for URTIs

Multiple responses were allowed in this variable. Cough 236 (32.1%), running nose 114 (19.4%), fever (temperature >38°C) 115 (15.6%), exudates in throats 84 (11.4%), chest pain 70 (9.5%), difficulty in breathing 38 (5.2%), and inflamed ear 26 (3.5%) constituted the most common clinical presentations which affected the physician's decision to prescribe antibiotics for URTIs

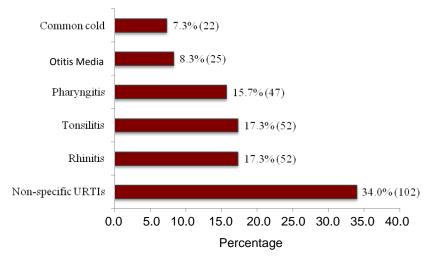


Figure 1. Type of URTIs diagnoses (N=300).

(Figure 2).

The multivariable logistic regression analysis revealed that five independent variables had a significant influence on the antibiotic prescription. In the adjusted analysis (additional adjustment such as sex and age) patients who had fever, chest pain and difficulty in breathing had reduced odds of being prescribed with antibiotic as compared to those without such symptoms (AOR=0.02, 95% CI: 0.008-0.09), (AOR=0.29, 95% CI: 0.12-0.74) and (AOR=0.25, 95% CI: 0.07-0.85), respectively. Moreover, patients who had cough and running nose had higher odds of being prescribed with antibiotic as compared to those without such symptoms (AOR=16.41, 95% CI: 1.95-138.19) and (AOR=1.98, 95% CI: 1.04-3.77), respectively (Table 3).

DISCUSSION

The objective of the present study was to assess physician prescription practice on antibiotics for URTIs in order to have baseline information on antibiotic use and thereafter make recommendations to stake holders. It has been documented that most URTIs are of viral origin in 80% of cases (Kunda et al., 2015), however, physicians in many settings frequently prescribe antibiotics for these illnesses and contributing to increasing antibiotic over-prescribing is a problem in many settings (Easton and Saxena, 2010; Kunda et al., 2015; Fletcher-Lartey et al., 2016; Zhang et al., 2017) and the present results show that Tanzania is not an exception.

In this study, it was found that a significant number of patients with URTIs were prescribed with at least one antibiotic (66.7%). This prevalence was notably high and

if not prevented it will continue to escalate the problem of antibiotic resistance. The present study, reports a relatively lower prevalence as compared to a studies by Gwimile et al. (2012) with a prevalence of 84.9%, a study in Namibia, with a prevalence of 78% among adults and children (Kunda et al., 2015) and in Malaysia, a prevalence of 68.4% (Teng et al., 2004). The reason for lower prevalence could be because of the availability of supportive environments such as laboratory facilities and effective hospital policies that influence antibiotic prescribing behaviors (Lyimo et al., 2018).

The most common URTIs diagnosed was non-specific URTIS (34.0%) followed by rhinitis and tonsillitis both 17.3%. This result was lower than studies conducted in Windhoek, Namibia and North Trinidad where it was 45and 54.5%, respectively (Mungrue et al., 2009; Kunda et al., 2015). This study shows that amoxicillin alone was the preferred drug for almost all URTIs. The same scenario was reported in Trinidad whereby amoxicillin alone or with clavulanate was the most frequently prescribed antibiotic for all URTIs (Mohan et al., 2004). This is of concern because prescribing antibiotics for these conditions in adults and children does not have any therapeutic benefits, but only increases the risk of developing antibiotic resistance. In addition, antibiotics do not warrant a better outcome in terms of cure or persistence of symptoms in patients who receive antibiotics compared to those who do not (Snellman et al., 2013).

Regarding signs and symptoms that influenced physician's decision to prescribe antibiotics for URTIs, fever (temperature >38°C), chest pain, cough, running nose and difficulty in breathing were the factors mostly affecting physician's decision to prescribe antibiotics. In this study, patients with fever, chest pain and difficulty in

Table 2. Antibiotics prescribed for URTIs.

					URTIs [n(%)]		
Antibiotic		Pharyngitis	Rhinitis	Tonsillitis	Common cold	Otitis media	Non-specific URTIs	Total
Are evicillie	No	24 (51.1)	32 (61.5)	36 (69.2)	6 (27.7)	23 (92.0)	84 (82.4)	206 (68.7)
Amoxicillin	Yes	23 (48.9)	20 (38.5)	16 (30.8)	16 (72.7)	2 (8.0)	18 (17.6)	94 (31.3)
Ampicillin	No	36 (76.6)	49 (94.2)	38 (73.1)	22 (100)	18 (72.0)	101 (99.0)	264 (88.0)
Ampicium	Yes	11 (23.4)	3 (5.8)	14 (26.9)	0 (0.0)	7 (28.0)	1 (1.0)	36 (12.0)
Co-amoxiclav	No	38 (80.9)	51 (98.1)	48 (92.3)	22 (100)	23 (92.0)	100 (980)	264 (88.0)
CO-amoxiciav	Yes	9 (19.1)	1 (1.9)	4 (7.7)	0 (0.0)	2 (8.0)	2 (2.0)	36 (12.0)
Cloxacillin	No	46 (97.9)	52 (100)	52 (100)	22 (100)	21 (84.0)	100 (980)	282 (94.0)
Cioxaciiiii	Yes	1 (2.1)	0 (0.0)	0 (0.0)	0 (0.0)	4 (16.0)	2 (2.0)	18 (6.0)
Ceftriaxone	No	46 (97.9)	52 (100)	43 (82.7)	22 (100)	16 (64.0)	101 (99.0)	293 (97.7)
Centraxone	Yes	1 (2.1)	0 (0.0)	9 (17.0)	0 (0.0)	9 (36.0)	1 (1.0)	7 (2.3)
Clarithromycin	No	43 (91.5)	50 (96.2)	46 (88.5)	22 (100)	25 (100)	102 (100)	280 (93.3)
Clantinoniyein	Yes	4 (8.5)	2 (3.8)	6 (11.5)	0 (0.0)	0 (0.0)	0 (0)	20 (6.7)
Ampiclox	No	43 (91.5)	52 (100)	47 (90.4)	21 (95.5)	15 (60.0)	100 (980)	289 (96.3)
Ampiciox	Yes	4 (8.5)	0 (0.0)	5 (9.6)	1 (4.5)	10 (40.0)	2 (2.0)	11 (3.7)
Chloramphenicol	No	4 5(95.7)	52 (100)	48 (92.3)	22 (100)	24 (96.0)	102 (100)	278(92.7)
Chioramphenicol	Yes	2 (4.3)	0 (0.0)	4 (7.7)	0 (0.0)	1 (4.0)	0 (0)	22(7.3)
	No	47 (100)	52 (100)	48 (92.3)	22 (100)	25 (100)	102 (100)	293 (97.7)
Penicillin G	Yes	0 (0.0)	0 (0.0)	4 (7.7)	0 (0.0)	0 (0.0)	0 (0)	7 (2.3)
Gentamicin	No	47(100)	52 (100)	52 (100)	22 (100)	21 (84.0)	102 (100)	296 (98.7)
	Yes	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	4 (16.0)	0 (0)	4(1.3)
Doutrovoline	No	46 (97.9)	47 (90.4)	51 (98.1)	20 (90.9)	25 (100)	99 (97.1)	288 (96.0)
Doxycycline	Yes	1 (2.1)	5 (9.6)	1 (1.9)	2 (9.1)	0 (0.0)	3 (2.9)	12 (4.0)
Azithromusia	No	47 (100)	52 (100)	50 (96.2)	22 (100)	21 (84.0)	102 (100)	294 (98.0)
Azithromycin	Yes	0 (0.0)	0 (0.0)	2 (3.8)	0 (0.0)	4 (16.0)	0 (0)	6 (2.0)
Any Antibiotic	No	0 (0.0)	14 (26.9)	23 (45.1)	9 (40.9)	9 (36.0)	20 (19.2)	100 (33.3)
Any Antibiotic	Yes	47 (100)	38 (73.1)	28 (54.9)	13 (59.1)	16 (64.0)	82 (80.4)	200 (66.7)

breathing were less likely to be prescribed with antibiotics. In other settings, patients with fever and cough were reported to be prescribed with antibiotics assuming that they were more severely ill (Akkerman et al., 2005). In areas like Tanzania where malaria is common, symptoms of fever and cough may also be shared with those of malaria. In such scenario, it is likely that clients who presented with these symptoms may have been prescribed with antimalarial, and this has been evidenced in different studies conducted in Kilimanjaro region, Northern Tanzania (Hertz et al., 2012; Crump et al., 2013; Kajeguka et al., 2016, 2017).

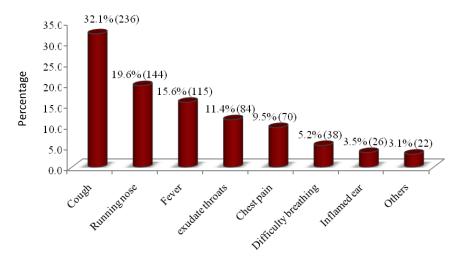


Figure 2. Signs and symptoms prompted antibiotic prescription (n=736, multiple responses were allowed).

Categories		Frequency [n (%)]	COR (95% CI)	AOR (95% CI)	^a AOR (95% CI)
Fever	Yes No	115 (38.3) 185 (61.7)	0.02 (0.008-0.08)	0.03 (0.009-0.10)	0.02 (0.008-0.09)
Chest pain	Yes No	70 (23.3) 230 (76.7)	0.26 (0.13-0.55)	0.24 (0.10-0.58)	0.29 (0.12-0.74)
Cough	Yes No	236 (78.7) 64 (21.3)	43.96 (5.99-322.39)	12.89 (1.60-103.59)	16.41 (1.95-138.19)
Running nose	Yes No	114 (48.0) 156 (52.0)	2.87 (1.73-4.74)	2.04 (1.08-3.86)	1.98 (1.04-3.77)
Exudates in throat	Yes No	84 (28.0) 216 (72.0)	0.89 (0.53-1.49)	-	-
Inflamed ear	Yes No	26 (8.7) 274 (91.3)	2.76 (0.60-12.72)	-	-
Difficulty in breathing	Yes No	38 (12.7) 262 (87.3)	0.31 (0.14-0.70)	0.28 (0.10-0.74)	0.25 (0.07-0.85)

Table 3. Signs and symptoms and antibiotic prescribed for URTIs.

COR: Crude odds ratio, AOR: adjusted odds ratio. Only variables that were predictors of antibiotic prescription (set in the bivariate analysis to p<0.05) were included in multivariate analysis. ^aIncludes additional adjustment such as sex and age.

The CDC and WHO recommend a performance of a group A β -hemolytic streptococci test prior to treatment of sore throat with antibiotics (Biezen et al., 2015). Laboratory test (such as culture and sensitivity) was

obtained for investigation for about 136 (47.1%) patients diagnosed with URTIs before antibiotic therapy was prescribed. This indicates that most prescribers are still not well aware of the concept and implications of antibiotic resistance. Therefore, they do not take laboratory tests into consideration when they decide whether or not to prescribe antibiotics for URTIs.

In order to reduce the irrational use of antibiotic prescribing in URTIs, it is of greatest importance that at the health facility level, the health care providers are trained on appropriate antibiotic prescription. This will assist in alleviating antibiotic overprescribing and the consequence of antibiotic resistance. Health workers' continuing education should be strengthened through conferences and seminars. Equally important is the periodic antibiotic use review, which can provide feedback to prescribers in health facilities on antibiotic use expenditure and resistance patterns. Also, there is obvious need for an antibiotic stewardship committee that will follow-up closely and enable care providers to rationally prescribe antibiotics.

Additionally, Tanzanian Ministry of Health, Community Development, Gender, Seniors and Children should consider adopting a strategy of delayed prescription or delayed antibiotic use, which has shown to be effective in reducing antibiotic usage for URTIs (Spurling et al., 2013; Ryves et al., 2016). In a Cochrane Review it has been highlighted that delayed prescribing may be a suitable compromise in place of immediate prescribing to significantly reduce unnecessary antibiotic use and thereby reduce antibiotic resistance while maintaining patient safety and satisfaction levels (Spurling et al., 2017). Moreover, regular training in antibiotic management for healthcare professionals is paramount, also there should be an awareness regarding antibiotics among patients and the community (Mbwambo et al., 2017). Further research is recommended to identify factors contributing to antibiotic over-prescribing in URTIs in Tanzania, which will also identify barriers of compliance to standard treatment guideline.

STRENGTHS AND LIMITATIONS

The study utilized systematic random sampling of prescriptions in KCMC and captured a broad view of antibiotic prescribing for URTIs in this setting. The study was conducted at one health facility. This prevented generalization of the results to the larger population of Tanzania. This study relied on the diagnosis written on the prescription and clinical presentation of the patient not the microbiological culture, which lead to a failure to comment on the appropriateness of the antibiotic prescription in relation to laboratory results.

CONCLUSION

In URTI treatment, use of antibiotic is always an area of concern. An irrational prescription practice of antibiotics is an important public health issue that affects the community. Use of antibiotic for the treatment of URTIs is evidently inappropriate unless the infection was proven to be bacteria.

CONFLICT OF INTERESTS

The authors have not declared any conflict of interests

REFERENCES

- Akkerman AE, Kuyvenhoven MM, Wouden JC, van der VTJM (2005). Determinants of antibiotic overprescribing in respiratory tract infections in general practice. Journal of Antimicrobial Chemotherapy 56:930-936.
- Al-Enezi AK, Fiala LE, Abu-Zaid LZ, Alsolami S, Taha A, Awwad K (2011). Determinants of Antibiotics Prescribing for Upper Respiratory Tract Infections among Primary Health Care Physicians in Al-Khober Area, Saudi Arabia. Medical Journal Cairo University 79:99-104.
- Biezen R, Pollack AJ, Harrison C, Brijnath B, Grando D, Britt HC (2015). Respiratory tract infections among children younger than 5 years: current management in Australian general practice. The Medical Journal of Australia 202:262-265.
- Bogaert D, Groot R, De Hermans PWM (2004). Streptococcus pneumoniae colonisation: The key to pneumococcal disease. Lancet Infectious Diseases 4:144-154.
- Centre for Diseases Control (CDC) (2017). Adult Treatment Recommendations: Community: Antibiotic Use [WWW Document]. Centre for Diseases Control and Prevention. URLhttps://www.cdc.gov/antibiotic-use/community/for-hcp/outpatienthcp/adult-treatment-rec.html [accessed on 2017].
- Centre for Clinical Practice (2008). Respiratory Tract Infections -Antibiotic Prescribing: Prescribing of antibiotics for self-limiting respiratory tract infections in adults and children in primary care [WWW Document]. National Institute for Health and Clinical Excellence: NICE clinical guideline 69. URL https://www.ncbi.nlm.nih.gov/pubmedhealth/PMH0010014/pdf/PubMe dHealth_PMH0010014.pdf [accessed on 2008].
- Chaw PS, Schlinkmann KM, Raupach-Rosin H, Karch A, Pletz MW, Huebner J (2018). Antibiotic use on paediatric inpatients in a teaching hospital in the Gambia, a retrospective study. Antimicrobial Resistance and Infection Control 7(1):82.
- Chilongola J, Msoka E, Juma A, Kajeguka DCDC, Semvua H, Semuva H (2015). Antibiotics prescription practices for provisional malaria cases in three hospitals in Moshi, northern Tanzania. Tanzania Journal of Health Research 17(3):1-10.
- Chonmaitree T, Revai K, Grady JJ, Clos A, Patel JA, Nair S, (2008). Viral Upper Respiratory Tract Infection and Otitis Media Complication in Young Children. Clinical Infectious Diseases 46:815-823.
- Cotton M, Innes S, Jaspan H, Madide A, Rabie H (2008). Management of upper respiratory tract infections in children. South African family practice: official journal of the South African Academy of Family Practice/Primary Care 50:6-12.
- Crump A, Morrissey A, Nicholson W, Massung R, Stoddard R, Galloway RL (2013). Etiology of severe non-malaria febrile illness in Northern Tanzania: a prospective cohort study. PLoS Neglected Tropical Diseases 7:1-9.
- Easton G, Saxena S (2010). Antibiotic prescribing for upper respiratory tract infections in children: How can we improve? London Journal of Primary Care 3(1):37-41.
- Faden H, Duffy L, Wasielewski R, Wolf J, Krystofik D, Tung Y (1997). Relationship between Nasopharyngeal Colonization and the Development of Otitis Media in Children. Journal of Infectious Diseases 175(6):1440-1445.
- Fletcher-Lartey S, Yee M, Gaarslev C, Khan R (2016). Why do general practitioners prescribe antibiotics for upper respiratory tract infections to meet patient expectations: A mixed methods study. BMJ Open 6:1-8.
- Givon-lavi N, Fraser D, Porat N (2002). Spread of *Streptococcus* pneumoniae and Antibiotic-Resistant *S. pneumoniae* from Day-Care

Center Attendees to Their Younger Siblings. Journal of Infectious Diseases 186:1608-1614.

- Gwimile JJ, Shekalaghe SA, Kapanda GN, Elton Richard K (2012). Antibiotic prescribing practice in management of cough and/or diarrhoea in Moshi Municipality, Northern Tanzania: cross-sectional descriptive study. Pan African Medical Journal 12:1-8.
- Haque M (2017). Antimicrobial Use, Prescribing, and Resistance in Selected Ten Selected Developing Countries: a Brief Overview. Asian Journal of Pharmaceutical and Clinical Research 10(8): 37–45.
- Hertz JT, Munishi M, Ooi EE, Howe S, Lim WY, Chow A (2012). Chikungunya and dengue fever among hospitalized febrile patients in northern Tanzania. The American Journal of Tropical Medicine and Hygiene 86(1):171-177.
- Kajeguka DC, Desrochers RE, Mwangi R, Mgabo MR, Alifrangis M, Kavishe RA (2017). Knowledge and practice regarding dengue and chikungunya: a cross-sectional study among Healthcare workers and community in Northern Tanzania. Tropical Medicine and International Health 22(5):583-593.
- Kajeguka DC, Kaaya RD, Mwakalinga S, Ndossi R, Ndaro A, Chilongola JO (2016). Prevalence of dengue and chikungunya virus infections in north-eastern Tanzania: a cross sectional study among participants presenting with malaria-like symptoms. BMC Infectious Diseases 16(1):183.
- Kunda M, Haoses-Gorases L, Goraseb M (2015). An Investigation of Antibiotic Prescribing in Patients with Upper Respiratory Tract Infections (Urtis) at Katutura Health Centre, Windhoek, Namibia. Single Cell Biology 4:1-7.
- Levy-Hara G, Amábile-Cuevas CF, Gould I, Hutchinson J, Abbo L, Saxynger L (2011). "Ten commandments" for the appropriate use of antibiotics by the practicing physician in an outpatient setting. Frontiers in Microbiology 2:1-7.
- Llor C, Moragas A, Bayona C, Cots JM, Molero JM, Ribas J (2017). The STOP-AB trial protocol: Efficacy and safety of discontinuing patient antibiotic treatment when physicians no longer consider it necessary. BMJ Open 7:1-7.
- Lyimo S, Geoffrey S, Emidi B, Mgabo MR, Kajeguka DC (2018). Crosssectional Survey on Antibiotic Prescription Practices Among Health Care Providers in Rombo District, Northern Tanzania. East African Health Research Journal 2:1-8.
- MacIntyre CR, Chughtai AA, Zhang Y, Seale H, Yang P, Chen J (2017). Viral and b.acterial upper respiratory tract infection in hospital health care workers over time and association with symptoms. BMC Infectious Diseases 17(1):553.
- Mbwambo G, Emidi B, Mgabo M, Sigalla G, Kajeguka D (2017). Community knowledge and attitudes on antibiotic use in Moshi Urban, Northern Tanzania: Findings from a cross sectional study. African Journal of Microbiology Research 11(25):1018-1026.
- Mohan S, Dharamraj K, Dindial R, Mathur D, Parmasad V, Ramdhanie J (2004). Physician behaviour for antimicrobial prescribing for paediatric upper respiratory tract infections: A survey in general practice in Trinidad, West Indies. Annals of Clinical Microbiology and Antimicrobials 3(1):11
- Mungrue K, Brown T, Hayes I, Ramroop S, Thurston P, Pinto PL (2009). Drugs in upper respiratory tract infections inpaediatric patients in North Trinidad. Pharmacy practice 7:29-33.
- Ouédraogo S, Traoré B, Bi ZABN, Yonli FT, Kima D, Bonané P (2014). Viral etiology of respiratory tract infections in children at the pediatric hospital in Ouagadougou (Burkina Faso). PLoS ONE 9:1-7.
- Peroš-Golubičić T, Tekavec-Trkanjec J (2015). Upper respiratory tract infections. In Textbook of Respiratory and Critical Care Infections. Jaypee Brothers Medical Publishers(P) Ltd. pp. 40-43.

- Ryves R, Eyles C, Moore M, McDermott L, Little P, Leydon GM (2016). Understanding the delayed prescribing of antibiotics for respiratory tract infection in primary care: A qualitative analysis. BMJ Open 6(11):e011882.
- Simoes EAF, Cherian T, Chow J, Shahid-Salles S, Laxminarayan R, John TJ (2006) Acute Respiratory Infections in Children. In Disease Control Priorities in Developing Countries (ed. by Jamison DT, Breman JG, Measham AR, et al.). pp. 483-497.
- Snellman L, Adams W, Anderson G, Godfrey A, Gravley A, Johnson K (2013). Health Care Guideline: Diagnosis and Treatment of Respiratory Illness in Children and Adults [WWW Document]. Institute for Clinical Systems Improvement. URL https://www.sunshinehealth.com/content/dam/centene/Sunshine/pdfs /2_respillness.pdf [accessed on 2013].
- Spurling GKP, Mar CB, Del Dooley L, Foxlee R, Farley R (2013). Delayed antibiotics for respiratory infections. Cochrane Database of Systematic Reviews.
- Spurling GKP, Mar CB, Del DL, Foxlee R, Farley R (2017). Delayed antibiotic prescriptions for respiratory infections. Cochrane Database of Systematic Reviews.
- Symekher SML, Ochieng W, Simwa J (2009). Prevalence of viral aetiologies in children with acute respiratory infections in Nairobi, Kenya. Tanzania Journal of Health Research 11:90-93.
- Teng CL, Leong KC, Aljunid SM, Cheah M (2004). Antibiotic Prescription In Upper Respiratory Tract Infections. Asian Pacific Family Medicine 3:38-45.
- Yoon YK, Park C, Kim JW, Hwang K, Lee SY, Kim TH (2017). Guidelines for the Antibiotic Use in Adults with Acute Upper Respiratory Tract Infections. Infection and Chemotherapy 49:326-352.
- Zhang Z, Hu Y, Zou G, Lin M, Zeng J, Deng S (2017). Antibiotic prescribing for upper respiratory infections among children in rural China: A cross-sectional study of outpatient prescriptions. Global Health Action 10(1):1-8.

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Full Length Research Paper

Simple bioanalytical method development and validation of micronised Domperidone 20 mg tablets using LCMS-MS and its pharmacokinetic application in Healthy Indian Volunteers

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The current investigation deals with an validated Liquid chromatography-mass spectrometry/mass spectrometry (LC-MS/MS) analytical method for the quantification of micronised domperidone in plasma of human volunteers. The validation of LC-MS/MS method was accomplished by evaluating the inter-day and intra-day precision and accuracy in a linear concentration range of 3.33-100 ng/ml. The entire study was an attempt to evaluate the comparison index of the bioavailability study of micronised domperidone tablet formulation with that of conventional domperidone tablet containing 20 mg of domperidone. Both the formulations were given orally as a single dose cross over design. The washout period was taken as 1 week. A single-dose, two-sequence, two-treatment, two-period crossover Bioequivalence study of two formulation were performed on 12 Indian healthy male volunteer. The estimation of domperidone concentration in human plasma was determined by the validated LC-MS/MS method. The various pharmacokinetics parameters like peak plasma concentration (C_{max}), and time to reach peak plasma concentration (t_{max}), area under the plasma concentration-time curve (AUC_{0-t}), area under the plasma concentration-time curve from zero to infinity (AUC_{0-∞}), of both the formulations were evaluated and compared. The results evaluated by estimated pharmacokinetic parameters did not find any statistically significant difference between the two formulations. The relative bioavailability of micronized test formulation was found to be 104.62% to that of reference conventional formulation.

Key words: Bioequivalence, domperidone, liquid chromatography-mass spectrometry/mass spectrometry (LC-MS/MS) analysis, pharmacokinetics.

INTRODUCTION

Gastroesophageal reflux disease (GERD) can be defined as one

as one of the most common incident related to upset in

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Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u> the gastro intestinal system (Hosseini et al., 2017; Dent et al., 2005). The recent advancement of medical sciences has already explored several options to treat and manage GERD effectively. Among them Acid suppression is one of the best treatment strategy to counter GERD symptoms for this reasons, Acid suppressing agents proton pump inhibitors (PPIs) provides the rapid and smooth symptomatic relief and heals esophagitis in a high proportion of patients (Dent et al., 2005). Domperidone can be considered as one of the most effective antiemetic drug for the treatment of mild to severe GERD symptoms Domperidone has dual mechanism of action. Domperidone can act as prokinetic agent which can stimulate the sphincter muscle of duodenam and easily can induce the effect of prokinetic movement. In addition, domperidone can block D₂ receptor antagonist in chemo trigger receptor zone (CTZ). The problem associated with domperidone for the treatment of GERD is its less bioavailability Domperidone is a water insoluble drug, domperidone is one of the ideal candidate to increase the solubility and as well as bioavailability. Here we developed a sustained release formulation with micronised domperidone. The main objective of our research work is to access the bioavailability of sustained release formulation and compare it with conventional non micronised formulation. Bioequivalent study of water insoluble drug has been executed by several researchers but very few related to micronised formulation has been addressed in a proper scientific way. Thorough literature survey finds that several methods was performed for conducting bioequivalent study but an attempt was made here to perform bioequivalent study of micronised domperidoene formulation as compare to conventional domperidone dosage form (Toyama et al., 2015; Bhadoriya et al., 2018; Blandizzi et al., 2015; Censi et al., 2015; Benet, 2013: Rockville, 2001).The interesting research conclusion which may come out from this study can be described as whether micronised drugs formulated as sustained release matrix tablet dosage form are able to deliver better and sustained bioavailability.

MATERIALS AND METHODS

Chemicals reagents and drug product

Raw domperidone (API) was provided as gift samples as by Kusum Healthcare, Punjab, India. HPLC grade methanol and Ethyl acetate were procured from Merck India Pvt. Ltd. (Mumbai). Milli Q water purification system was installed to acquire High Performance Liquid Chromatography (HPLC) grade water. The human blank plasma sample with EDTA-K₃ anticoagulant was procured from Bioequivalence Study Centre, Jadavpur University, Kolkata, India. Test product: Tablet containing micronized Domperidone 20 mg and Reference product: Domstal, from Torrent Pharmaceutical Ltd, (Torrent House, Ahmedabad, India), containing domperidone 20 mg.

Instrumentation

The LC system was purchased from Shimadzu (Kyoto, Japan). API 2000 triple quadrupole mass spectrometer (MDS Sciex, Canada) with electrospray ionization (ESI) source was used for detection of the compound. Data acquisition was done with Analyst 1.4.1. software. Chromatographic separation was performed on a standard C8 column, 50 mm × 3 mm, 3 µm i.d (Phenomenex, USA).

Products studied

The following test and reference products were used in the present study. Test product: Tablet containing micronized domperidone 20 mg and reference product: Domstal, from Torrent Pharmaceutical Ltd, (Torrent House, Ahmedabad, India), containing domperidone 20 mg.

Chromatographic conditions

The entire chromatographic analysis was executed at ambient atmospheric temperature with a runtime 5 min. The injection volume was taken as 20 μ l. The composition of water: methanol (2: 98, v/v) was used as mobile phase containing 0.5% formic acid with a flow rate of 1 ml min⁻¹. The column oven was kept at 23°C. While the temperature of auto sampler was maintained at 10°C. The mass spectra of the compounds were acquired by using Electrospray ionization (ESI) with multiple reactions monitoring (MRM) technique. The entire ionization of the drug was accomplished in positive ionization mode. The important tuning parameters were calculated and optimized by injecting 100 ng mL⁻¹ of standard solution containing all two drugs including internal standard. The validation parameters like sensitivity, accuracy, precision, stability, recovery, reproducibility and system suitability were measured in accordance with the US-FDA bioanalytical method guidelines (Bhadoriya et al., 2018).

Study design

The whole bioequivalent study was executed under fasting conditions as a two-sequence two-period crossover study. The study design was based on free randomization. The drug was administered with single-dose. Between the two periods minimum one week of dosing interval was taken as a washout period (Blandizzi et al., 2015) prior to the study. The experimental protocol was reviewed and approved by Institutional Ethical Committee (ICE) of Jadavpur University, Kolkata, India. The volunteers were enrolled after thorough investigation including medical history, vital parameters of physical examination, laboratory investigation, drug screening, ECG and HIV/hepatitis status. An experience clinical pharmacologist was actively present throughout the study to guide and monitor the entire study.

Drug administration and blood sample collection

12 non-smokers healthy Indian male volunteers were screened for the study. The age of the volunteers were between 19 to 33 years (29 ± 3.49) with a standard body mass index between 19 to 27 (24.66 ± 3.76) . The whole study was executed under the regulation and guidance issued by U.S. Food and Drug Administration (FDA) and European Agency for the Evaluation of Medicinal Products (EAMP) from time to time (Censi et al., 2015; Benet, 2013). A pre planed blood sampling schedule was designed to evaluate the rate and extent of absorption in such a manner so that all the important

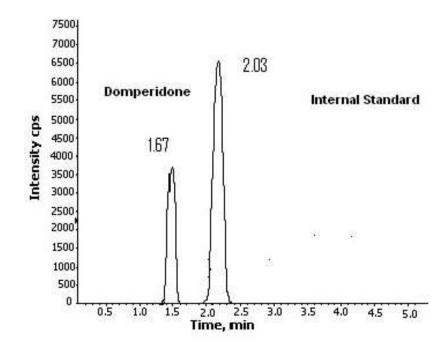


Figure 1. Retention time of Domperidone and internal standard.

pharmacokinetic parameters can be calculated properly (Rockville, 2001) Committee for Proprietary Medicinal Products(CPMP) (1991).

Total of 13 blood samples were collected from each volunteers at various interval of time including 0, 1.0, 2.0, 3.0, 4.0, 5.0, 6.0, 7.0, 8.0, 10.0, 12.0, 24.0 and 48.0 h in the sterile EDTA added centrifuge tubes. After the centrifugation the plasma was separated entirely and stored at a temperature of -20°C.

Pharmacokinetic analysis

Noncompermental pharmacokinetic model were used here to calculate various pharmacokinetic parameters of domperidone. The peak plasma drug concentration (C_{max}) and time to reach peak plasma concentration (t_{max}) were calculated directly from the results obtained after the analysis of the drug. The elimination half-life ($t_{1/2}$) was calculated by using the formula of 0.693/K_{e.}, where K_e is considered as apparent elimination rate constant calculated from the slope of the terminal log linear phase. Area calculation of trapezoidal rule was implemented to find AUC_{0-t.} AUC_{0-∞} was also estimated according to the following standard formula:

$$AUC_{0-\infty} = AUC_{0-t} + C_{last} / K_e,$$

where C_{last} is the last quantifiable plasma concentration (US Food and Drug Administration, 2017)

Statistical analysis

Pharmacokinetic parameters of each subject were studied thoroughly on the basis of statistical approach. Bioequivalence study parameters like AUC_{0-t} , $AUC_{0-\infty}$ and C_{max} values were compared as primary variables. Statistical tool of analysis of variance (ANOVA), including treatment, period and subject were

applied for these primary parameters and also for log-transformed values of these parameters. The statistical approach of bioequivalence analysis was done according to guidance of Committee for Proprietary Medicinal Products (Censi et al., 2015). The experimental test formulation was considered to be bioequivalent to reference formulation when 90% confidence interval (CI) for the ratio between each pharmacokinetic parameters of test and reference was found to be within the fixed equivalence range of 80-125% (Nation and Sansom, 1994).

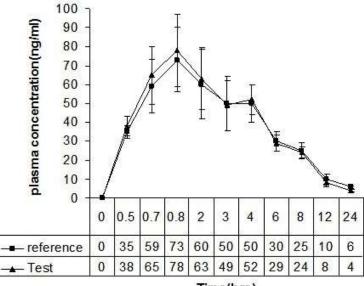
RESULTS

The developed bioanalytical method used for the estimation of domperidone in biological matrix was found to be accurate and sensitive. The peaks of domperidone and Internal standard both were found to be well resolved. No interference observed was in the chromatogram of blank plasma sample during. During LC-MS/MS analysis (Figure 1). The retention time (RT) of domperidone and Internal Standard was found to be at 1.67 and 2.03 min respectively. The lower limit of quantification (LLOQ) for domperidone in plasma was noted as 3.33 ng /ml. The peak area ratio (domperidone: Internal standard) between concentration and was found to be linear within the range of 3.33 ng/ml to 500 ng/ml $(r^2=0.9998)$. Stability, absolute recovery, within-day and between-day, precision and accuracy was estimated for three different quality control points at low, medium, and high levels (5, 50 and 80 ng/ml). Mean drug plasma concentration at various interval of time after oral

Parameter	Test	Reference	90% CI (Log-transformed data)
AUC _{0-t} (ng. h /ml)	510.55±24.25	488.05±33.52	0.99007 - 1.00296
AUC _{0-∞} (ng. h /ml)	563.69±31.25	545.98±36.95	0.98942 - 1.00080
C _{max} (ng/ml)	78.32±53.30	73.17± 51.55	0.99527 - 1.00836
t _{max} (h)	0.800±0.150	0.798±0.04	
K _e (h ⁻¹)	0.15±0.008	0.17±0.004	
t _{1/2} (h)	4.62±0.306	4.081±0.295	

Table 1. Mean (\pm SD, n = 12) pharmacokinetic parameters of 20 mg domperidone tablet for test and reference preparation.

 $AUC_{0-t_i} AUC_{0-e_i} C_{max_i} t_{max_i} K_{e_i} and t_{1/2} are the area under the plasma concentration-time curve upto 48h, area under the plasma concentration-time curve upto infinity, maximum plasma concentration, time to reach maximum plasma concentration, elimination rate constant, and half-life of a drug, respectively.$



Time(hrs)

Figure 2. Mean (\pm SD, n=12) plasma concentration-time profiles after administration of test and reference preparations in healthy Indian subjects [- \blacksquare - is test formulation graph and - \bullet - is reference formulation graph obtained by plotting time (h) on X-axis and plasma concentration (ng/ml) on Y-axis.

administration of reference and test products to healthy volunteers are depicted in Table 1. The comparison of all the major pharmacokinetic parameters for the drugs including ratios of C_{max} , AUC_{0-t}, and AUC_{0- ∞} were obtained within the range of 0.80-1.25 at 90% confidence interval.

DISCUSSION

The above described bioanalytical method used for estimation of domperidone in plasma matrix was found to be very simple, robust, accurate and sensitive. The entire therapeutic window was covered by the linearity range achieved for this assay (3.33 to 500 ng/ml). The peak of drug domperidone and Internal Standard were well resolved as shown in Figure 2. Throughout the whole experimental study, domperidone was found to be stable in biological matrixes. Final mean recovery of three different quality control sample for three freeze and thaw cycles was found to be 87.60% and coefficient of variation (CV) was noted as 4.26%.

The elimination half-life $(t_{1/2})$ of domperidone in various formulations was found to be in the range 4.21 to 5.02 h. For this reason, one-week wash out period was sufficient

between the two phases. Peak drug plasma concentration (t_{max}) was observed at 0.8 h after drug administration, and the last samples were sufficient for calculating at least 80% of AUC₀₋. After oral administration of reference drug the peak plasma concentration $C_{\mbox{\scriptsize max}}$ was found to be 73.17 \pm 21.55 ng/ml at the time 0.798 \pm 0.04 h (t_{max}). For the test preparation peak plasma concentration (C_{max}) was found to be 78.32±23.30 ng/ml at the time 0. 800±0.150 (t_{max}). AUC_{0-t} of the test and reference were found to be 510.55±24.25 ng h/ml versus 488.05±33.52 ng h /ml respectively and AUC_{0-∞} of the test and reference were found to be 563.69±31.25 ng h /ml versus 545.98±36.95 ng h /ml respectively. On the basis of calculation of comparison of the AUC_{0-t} for domperidone after single dose administration, the relative bioavailability of the test preparation was 105% to that of reference preparation.

The objective of the bioequivalence study is to confirm interchangeability between a test (innovator sample) and a generic drug (reference) formulation on the basis of efficacy and safety. When a pharmacological effect of certain drug is difficult to estimate, the plasma levels of a drug may be utilized as an indicator of clinical activity. For this reasons, domperidone plasma concentration obtained in this bioequivalent study suggest an equal clinical efficacy of the two brands tested and provide pharmacokinetic data from Indian healthy volunteers.

Conclusion

The 90% CI of C_{max} , AUC_{0-t} and $AUC_{0-\infty}$ of domperidone of these two preparations was found to be in acceptable range as mentioned earlier. There was no statistically significant difference for the treatment values. Both formulations were equal in terms of rate and extent of absorption. Consequently bioequivalence between two formulations can be concluded.

CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

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REFERENCES

- Benet LZ (2013).The role of BCS (Biopharmaceutics Classification System) and BDDCS (Biopharmaceutics Drug Disposition Classification System) in drug development. Journal of pharmaceutical sciences 102(1):34-42
- Bhadoriya A, Dasandi B, Parmar D, Shah PA, Shrivastav PS (2018). Quantitation of tadalafil in human plasma using a sensitive and rapid LC-MS/MS method for a bioequivalence study Journal of Pharmaceutical Analysis 8(4):271-276.
- Blandizzi C, Viscomi GC, Scarpignato C (2015). Impact of crystal polymorphism on the systemic bioavailability of rifaximin, an antibiotic acting locally in the gastrointestinal tract, in healthy volunteers. Drug design, development and therapy 9:1-11.
- Censi Ř, Rascioni R, di Martino P (2015). Changes in the solid state of anhydrous and hydrated formsof sodium naproxen under different grinding and environmental conditions: Evidence of theformation of new hydrated forms. European Journal of Pharmaceutics and Biopharmaceutics 92:192-203
- Committee for Proprietary Medicinal Products (CPMP) (1991). *Note for* Guidance: Investigation of Bioavailability and Bioequivalence. London: Working Party on the Efficacy of the Medicinal Products.
- Dent J, El-Serag H, Wallander MA, Johansson S (2005). Epidemiology of gastro-oesophageal reflux disease: a systematic review. Gut 54(5):710-717.
- Hosseini M, Salari R, Shariatmaghani S, Birjandi B, Salari M (2017). Gastrointestinal symptoms associated with gastroesophageal reflux disease, and their relapses after treatment with proton pump inhibitors: a systematic review. Electronic physician 9(6)4597- 4605.
- Nation RL, Sansom LN (1994) Bioequivalence requirements for generic products. Pharmacology and therapeutics 62(1-2):41-55.
- Ritschel WA, Drug Intelligenc. In: Handbook of Basic Pharmacokinetics. 4th ed. Hamilton: Hamilton Press; 1992. p.588.
- Rockville MD (2001) Guidance for industry, Bioanalytical Method Validation. US Department of Health and Human Services. Food and Drug Administration. Centre for Drug Evaluation and Research.
- Toyama K, Uchida N, Ishizuka H, Sambe T, Kobayashi S (2015). Single-dose evaluation of safety, tolerability and pharmacokinetics of newly formulated hydromorphone immediate-release and hydrophilic matrix extended-release tablets in healthy Japanese subjects without co-administration of an opioid antagonistThe Journal of Clinical Pharmacology 55(9):975-984
- US Food and Drug Administration (2017). Center for Drug Evaluation and Research. Guidance for industry: Dissolution testing of immediate release solid oral dosage forms. 2017, http://www.fda.gov/cder/Guidance/1713bp1.Pdf.

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African Journal of Pharmacy and Pharmacology

Full Length Research Paper

Synthesis, characterization, and anticancer activity against human breast cancer cell-line T47D studies of metal ion Cu(II) complex with 2,4,5-triphenylimidazole ligand

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The complex of metal ion Cu(II) with the ligand 2,4,5-trifenilimidazol has been successfully synthesized with mole ratio of metal and ligand 1:2 in N,N-dimethylformamide as a solvent. Complex synthesis results obtained green light crystalline solid. Complex absorbs UV-Vis light at 529 nm. Fourier transform infrared (FTIR) characterization results indicate occurrence of bonding of metals and ligand that is Cu-N in region 422.38 cm⁻¹. Results of elemental analyzer and Atomic Absorption Spectroscopy (AAS) analysis show the complex formed has the formula [Cu(L)₂(H₂O)₂].Cl₂. The molecular formula is also supported by the Thermal Gravimetric Analyzer data. Thermal Gravimetric Analyzer (TGA) analysis results showed that there was no water in the crystalline complex compounds. The cytotoxicity test complex compounds made by the method of 3-(4,5-dimetiltiazol-2-yl)-2,5-difeniltetrazolium bromide (MTT) and the IC₅₀ value of complex obtained 72.139 μ g/ml.

Key words: Copper(II), 2,4,5-trifenilimidazol, complex compound, characterization, anticancer.

INTRODUCTION

Complex compounds continue to be developed into useful tools in the field of medicine. Some synthesized compounds exhibit good activity in the field of medicinal chemistry. Cisplatin has been proven as a very effective chemotherapy agent to treat various types of cancer (Reedijk and Lohman, 1985). However, these platinumbased complex compounds cause side effects at certain doses and provide drug resistance during the therapeutic process (Renny et al., 2013). This led to the development of the discovery of new non-platinum-based complex

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Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u> compounds, in the hope of improving pharmacological properties, reducing side-effects, and obtaining different drug-specific targets (Qiao et al., 2011). Some of the transition metals used in the synthesis of anticancer complex compounds include Co(II), Ni(II), Cu(II), Pd(II), Ru(II) and Pt(II) (Budzisz et al., 2009; Ali et al., 2013).

Copper(II) metal is an essential element and plays an important role in the biological system of the human body, as a constituent of redox and hemocyanin enzymes (Linder and Maryam, 1996). As previously reported, complexes synthesized from Cu(II) metal ions with 2-(4'thiazolyl)benzimidazole and 2-(2-pyridyl)benzimidazole ligands have a cytotoxicity to liver cancer cells (hepatocellular carcinoma) (Devereux et al., 2007).

Ligands commonly used and continuously developed in studies of anticancer drug compounds have bound atoms of nitrogen and oxygen atoms, including derivatives of imidazole, benzamide, pyridine, and pyrazole (Goncalves et al., 2013; El Boraey, 2012; Tiwari et al., 2011; Budzisz et al., 2009). N-containing aromatic ligands such as pyridine, imidazole, and their derivatives (which are as electron donors similar to purine and pyrimidine bases) have been reported to possess in vitro anticancer properties such as Cisplatin (Deegan et al., 2006). The imidazole-based complex compounds showed anticancer activity against SK-MEL-31 skin cancer cells and tongue cancer cells CAL-27. The imidazole-based compounds also show cytotoxic effects on HepG2 liver cancer cells and A-498 bowel cancer, MCF-7 breast cancer, cervical cancer HeLa, and HL-60 blood cancers (Devereux et al., 2004; Bhat et al., 2011). Therefore, in this study, synthesized complex compounds of Cu (II) metal ion with 2,4,5-trifenylimidazole ligand and tested anticancer activity by MTT method in vitro assay against breast cancer cell T74D.

MATERIALS AND METHODS

The materials used in this study were copper(II)chloride dihydrate (CuCl₂.2H₂O) (Merck 99.0%), N,N-dimethylformamide (DMF) (Merck 99.8%), dimethyl sulfoxide (DMSO) (Merck 99.8%), 2,4,5-triphenylimidazole (Sigma-Aldrich 90%), methanol (Sigma-Aldrich 98%), breast cancer cell T74D (CVCL_0553), RPMI 1640 Medium (Gibco), Phospate-buffered saline 1X (PBS 1X) (Gibco), and Thiazoyl blue tetrazolium bromide (MTT) (Bio Basic).

Determination of maximum wavelength of $[Cu(L)_2(H_2O)_2].Cl_2$

The wavelength of the Cu(II) complex compound with 2,4,5-trifenyl imidazole was determined by continuous variation method. Continuous variation begins with a 1 mole molecular synthesis of CuCl₂.2H₂O and 2,4,5-triphenylimidazole, 1 mole with a volume ratio of 0:10, 1:9, 3:7, 5:5, 7:3, 9:1, and 10:1. Each of these solutions is in DMF and heated 3 h at 120°C. The formed solid is decanted and dried. The solid was dissolved in DMSO and measured the maximum wavelength (λ_{max}) with a UV-Vis

spectrophotometer, then graphed between absorbance as ordinate and mole fraction of metal as abscess.

Synthesis and characterization of [Cu(L)₂(H₂O)₂].Cl₂

The synthesis of these complex compounds was performed by the mole of metal and ligand 1:2. Watched the copper(II)chloride dihydrate and 2,4,5-triphenylimidazole in DMF.¹⁵ (Han et al., 2012). The complex solution was put into a vial, distilled for 30 min and heated at 120°C for 3 h. The mixture was then cooled to room temperature in vial covered with aluminium foil which has been given several small holes and left for 7 days to form solids and every day was washed with methanol to remove impurities contained in the mixture. The formed solid is decanted and dried. Subsequently, was characterized by UV-Vis Spectrophotometer, FTIR Spectroscopy, Atomic Absorption Spectroscopy (AAS), Thermal Gravimetric Analyzer (TGA), and CHN analyzer.

Anticancer activity

The breast cancer cell T74D with a density of 5×10^3 cells/well was distributed into 96 wells plate, incubated for 24 h at a 37°C CO₂ to attach. The medium was then replaced with fresh complete medium containing DMSO 0.1% (control), compound complex at concentration of 50, 25, 12.5, 6.25, 3.13, and 1.56 µg/ml, and incubated for 20 h (37°C/CO₂). Then into each well was added 100 µl RPMI containing MTT reagent and the plates incubated for an additional 4 h. Living cells react with MTT to form formazan crystals (Mosmann, 1983). After 4 h, the medium containing MTT was discarded and then added 50 µl DMSO solutions to dissolve the formazan crystals, homogenized on top of shaker for 10 min, then read with Microplate reader at wavelength 595 nm.

RESULTS AND DISCUSSION

Maximum wavelength of [Cu(L)₂(H₂O)₂].Cl₂

The solids of $CuCl_2.2H_2O$ and 2,4,5-triphenylimidazole with a 1:2 mole ratio reacted with DMF solution and heated for 3 h at 120°C. The complexes obtained are light green solids (72.127% of yield) and maximum wavelength complex with UV-VIS Spectrophotometer at 529 nm as shown in Figure 1. The result of 10x magnification photograph shows that the obtained solid is in the form of a needle as shown in Figure 2.

Analysis of functional groups with FTIR Spectroscopy

Characterization using FTIR spectroscopy was used to determine the presence of functional groups in complex and new bonds formed between metal and ligand. Comparison of FTIR spectrum of ligand and complex is as shown in Figure 3.

Based on the obtained data, the appearance of a new peak at a wavelength 422.38 cm⁻¹ can be seen. The peak is observed as a vibration of metals and ligands. This is in line with previous research showing that new peaks of

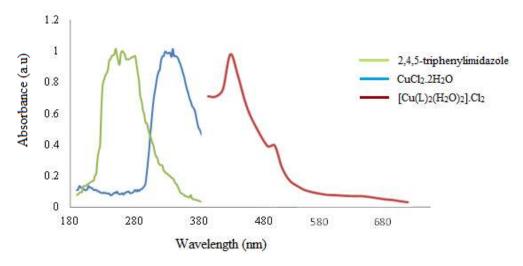


Figure 1. Maximum wavelength spectrum of CuCl₂.2H₂O (blue light), $[Cu(L)_2(H_2O)_2].Cl_2$ (red), and 2,4,5-triphenylimidazole ligand (green light). There was a maximum wavelength shift from the 2,4,5-triphenylimidazole ligand to the $[Cu(L)_2(H_2O)_2].Cl_2$ complex of 243 nm to 529 nm, and the maximum wavelength of CuCl₂.2H₂O at 338 nm. This shows that the complex $[Cu(L)_2(H_2O)_2].Cl_2$ has been formed.



Figure 2. The compounds of [Cu(L)₂(H₂O)₂].Cl₂ (right); 10x magnification (left).

complexes (metal bonds and ligands (Cu-N)) appear on a wavelength of 453 cm⁻¹, whilst Cu metal bonds with H_2O ligands appear at the of 534.25 cm⁻¹ wavelength indicating the presence of Cu-O (Gomathi and Murugan, 2014).

This indicates that the N-H bond detected is a tertiary N-H. While the complexes appear to peak with weak intensity in the area of 3400.27 cm^{-1} ; this peak is a characteristic of secondary amines located in the 2,4,5-trifenylimidazole ligand (Marzouk et al., 2013). The formation of secondary amine peaks is due to the synthesized complex structure [Cu(II)-2,4,5-triphenylimidazole] possessing two moles of ligand, so that the intensity of the secondary NH bond is stronger and therefore it can be detected even when the intensity is weak. In the spectra of complex, located in an area above 3000 cm⁻¹, it is slightly larger than the ligand

because of the presence of a coordinated O-H bond in the water as a ligand (Ullah et al., 2016). The uptake that appears at 1600 to 1700 cm⁻¹ is a C=N vibration in a 2,4,5-triphenylimidazole ligand coordinate with Cu metal ions.

Elementals analysis

Determination of the molecular formula of the complex can be obtained through the theoretical calculation approach of the composition of the formation of the complex. In this case, a complex formed of one metal and two ligands with the formula molecule $[(H_2O)_xCu-(L)_2(CI)_y]$.Cl_y can be seen. The addition of Cl outside the complex serves to neutralize the complex charge, so the amount of Cl will adjust to its charge. The theoretical

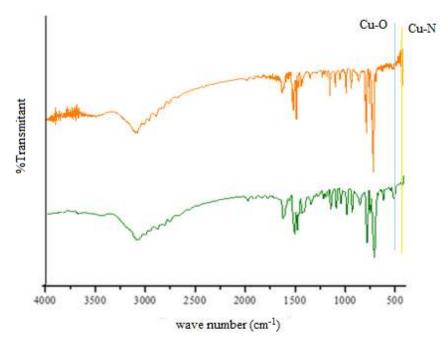


Figure 3. Comparison of FTIR spectrum of 2,4,5-triphenylimidazole ligand (orange) and complex compound [Cu(L)₂(H₂O)₂].Cl₂ (green).

approach is done by calculating the percentages of Cu, C, H, and N on the complex. Theoretically, the complex formed has the formula $[Cu(L)_2(H_2O)_2].Cl_2$ where two ligands bind one Cu metal according to continuous variation.

The result of the analysis shows that the synthesized complex contains Cu elements of 8.334%, C element of 66.2751%, H element 4.7284% and N element equal to 7.3861%. Based on the data, the percentage level of each element in theory from some complex structures that may be formed was calculated. The result of the calculation of elemental content theoretically approaching experimental measurement result is the prediction result of molecule formula of complex compound. Matches show results appropriate for the complex formula $[Cu(L)_2(H_2O)_2]$.Cl₂. These results indicate that the 2,4,5triphenylimidazole ligand only binds one Cu. The presence of a steric hindrance causes the 2,4,5triphenyilimidazole ligand to be very difficult to bind metals and can only bind one metal. This shows that H bound to N is not all replaced by Cu.

Thermal gravimetric analysis (TGA)

In the determination of complex molecular formulas, thermo gravimeters can provide specific information of a complex that decompose when it is heated. The complex thermo gravimetric analysis of Cu(II)-2,4,5-

triphenylimidazole was carried out at 25 to 600°C with a complex sample weight of 6.3670 mg. Based on the complex TGA curve of Figure 4, it can be seen that there is one stage of decomposition in the complex. Weight loss of 86.8791% occurring at 255.33 to 355.83°C indicates a complex decomposition consisting of 2 ligand molecules 2,4,5-triphenylimidazole, 2 molecules H₂O and 1 molecule Cl₂. This result corresponds to the theoretical weight that in weight reduction of 86.8791% is decomposition $((C_{21}H_{16}N_2)_2(H_2O)_2CI_2)$. The residue of 13.1209% (theoretically 8.333%) can be predicted as Cu, as in the previous study that CuO is the final residue of complex [Cu(6-hydroxypicolinate)₂(3-picolinate)₂] the (Kukovec et al., 2012). Weight loss does not occur at temperatures of 75 to 147°C. This means the complex does not contain crystalline water (Tamaekong et al., 2014).

Anticancer activity

In this research, cytotoxicity test was done by MTT method using T74D breast cancer cell. This test is used to determine the cytotoxic effect of tested compound. The classification of toxicity level of the extract based on IC_{50} , which is very high category (highly toxic) if it can kill 50% of cells at concentrations of 1 to 10 µg/ml, medium category (medium toxic) at concentrations of 10 to 100 µg/ml

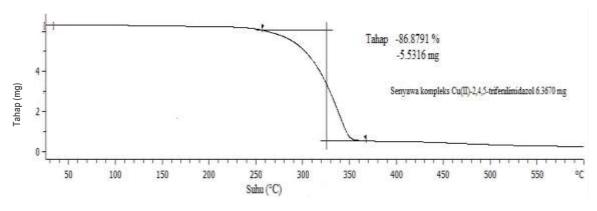


Figure 4. Thermal gravimetric analysis curve of [Cu(L)₂(H₂O)₂].Cl₂.

(Meyer et al., 1982). Based on the calculation, IC_{50} value for complex [Cu(L)₂(H₂O)₂].Cl₂ was obtained at 72.139 μ g/ml. Based on the IC₅₀, the [Cu(L)₂(H₂O)₂].Cl₂ complex belongs to the category of medium toxicity compound (medium toxic). This is in consistent with the previous research in which the CuL complex (L = 3-(1,3dioxoisoindolin-2-yl)-2,6-dioxopiperidine-1-carbodithioate) kills more cancer cells than its free ligands (Ali et al., 2013). The result of IC_{50} of $[Cu(L)_2(H_2O)_2].Cl_2$ complex in this study was greater than in previous study, where the assay performed the MTT was on complex [Cu(TBZH)₂(BZA)]-(BZA).0.5TBZH.H₂O wherein the TBZH ligand was 2-(4'-thiazole)benzimidazole and the BZA ligand was benzoic acid, IC₅₀ of 32 µM (Devereux et al., 2007).

CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

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REFERENCES

- Ali I, Wani AW, Saleem K, Hseih M (2013). Design and Synthesis of Thaliomide Based Dithiocarbamate Cu(II), Ni(II) and Ru(III) Complexes as Anticancer Agents. Polyhedron 56:134-143
- Bhat SS, Kumbhar AA, Heptullah H, Khan AA, Gobre VV, Geiji SP, Puranik VG (2011). Synthesis, Electronic Structure, DNA and Protein Binding, DNA Cleavage, and Anticancer Activi of Fluorophore-

Labeled Copper(II) Complexes. Inorganic Chemistry 50:545-558. Budzisz E, Magdalena M, Ingo-Peter L, Peter M, Urszula K, Marek R (2009). Synthesis and X-ray Structure of Platinum(II), Palladium(II) and Copper(II) Complexes with Pyridine–Pyrazole Ligands: Influence of Ligands' Structure on Cytotoxic Activity. *Polyhedron* 28:637–645.

- Deegan C, Coyle B, McCann M, Devereux M, Egan DA (2006). In vitro Anti-tumor Effect of 1,10-phenanthroline-5,6-dione(phendione), [Cu(phendone)₃](ClO₄)₂.4H₂O and [Ag(phendione)₂]ClO₄ using Human Epither Cell Lines. Chemico-Biological Interactions 164:115-125.
- Devereux M, McCann M, Shea DO, Kelly R, Egan D, Deegan C, Kavanagh K, McKee V, Finn G, (2004). Synthesis, Antimicrobial Activity and Chemotherapeutic Potential of Inorganic Derivatives of 2-(4'-thiazolyl)benzimidazole{thiabendazole}: X-ray Crystal Structure of [Cu(TBZH)₂Cl]Cl.H₂O.EtOH and TBZH₂NO₃ (TBZH = thiabendazole). Journal of Inorganic Biochemistry 98:1023-1031.
- Devereux M, Shea DO, Kellett A, McCann M, Walsh M, Egan D, Deegan C, Kedziora K, Rosair G, Muller-Bunz H (2007). Synthesis, X-ray Crystal Structures and Biomimetic and Anticancer Activities of Novel Copper(II)benzoate complexes incorporating 2-(4'thiazolyl)benzimidazole (thiabenzole), 2-(2-pyridyl)benzimidazole and 1,10-phenanthroline as Chelating Nitrogen Donor Ligands . Journal of Inorganic Biochemistry 101:881-892.
- El Boraey HA (2012). Coordination Behavior of Tetraaza [N4] Ligand towards Co(II), Ni(II), Cu(II), Cu(I) and Pd(II) Complexes: Synthesis, Spectroscopic Characterization and Anticancer Activity. Spectrochim Acta A. 97:255-262.
- Gomathi R, Ramu A, Murugan A (2014). Evaluation of DNA Binding, Cleavage, and Cytotoxic Activity of Cu(II), Co(II), and Ni(II) Schiff Base Complexes of 1-Phenylindoline-2,3-dione with Isonicatinohydrazide. Bioinorganic Chemistry and Applications 2014:1-13.
- Goncalves AC, Morias TS, Robalo MP, Maques F, Avecilla F, Matos CP, Santos I, Tomaz AI, Garcia MH (2013). Important Cytotoxicity of Novel Iron(II) cyclopentadienyl Complexes with Imidazole Based Ligands. Journal of Inorganic Biochemistry 129:1-8.
- Han S, Lough AJ, Kim JC (2012). Synthesis, Crystal Structures and Properties of Macrocyclic Copper(II) Compexes Containing Imidazole Pendants. Bulletin of the Korean Chemical Society 33:2381-2384.
- Kukovec B, Kaksa M, Popovic Z (2012). Synthesis and Characterization of a Copper(II) Complex with 6-hydroxypicolinic Acid and 3-Picoline. Croatica Chemica Acta 85:479-483
- Linder MC, Maryam HA (1996). Copper Biochemistry and Molecular Biology. American Journal of Clinical Nutrition 63:797-811.
- Marzouk AA, Abbasov VM, Talybov AH, Mohamed SK (2013). Synthesis of 2,4,5-Triphenyl Imidazole Derivatives Using Diethyl Ammonium Hydrogen Phospate as Green, Fast and Reusable Catalyst. World Journal of Organic Chemistry 1:6-10.

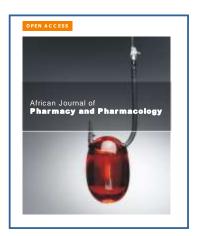
- Meyer BN, Ferrighi NR, Putnam JE, Jacobsen LB, Nichols DE, Mclaughlin JL (1982). Brine Shrimp:A Convenient General Bioassay for active Plant Constituents. Planta Medica 45:31-34.
- Mosmann T (1983). Rapid Colorimetric Assay for Cellular Growth and Survival: Application to Proliferation and Cytotoxicity Assays. Journal of Immunological Methods 65:55-63.
- Qiao X, Zhong YM, Cheng ZX, Fei X, Yan WZ, Jing YX, Zhao YQ, Jian SL, Gong JC, Shi-Ping Y (2011). Study of Potential Antitumor Mechanism of A Novel Schiff Base Copper(II) Complex:Synthesis, Crystal Structure, DNA Binding, Cytotoxicity and Apoptosis Induction Activity. Journal of Inorganic Biochemistry 105:728-737.
- Reedijk J, Lohman PHM (1985). Cis-platinum; synthesis, antitumor activity and mechanisms of action. Pharmaceuticals Weekbl Science 7:173-180.
- Renny SJ, Tomasevich L, Tallmadge H, Cullom D (2013). Method of Continuous Variations: Application Job Plot to the Study of Molecular Associations Organometallic Chemistry. Angewandte Chemistry International 52:2-18.

- Tamaekong N, Liewhiran C, Phanichphant S (2014). Synthesis of Thermally Spherical CuO Nanoparticles. Journal of Nanomaterials 2014:1-5.
- Tiwari AD, Mishrab AK, Mishraa SB, Mambaa BB, Majic B, Bhattacharya S (2011). Synthesis and DNA Binding Studies of Ni(II), Co(II), Cu(II) and Zn(II) Metal complexes of N1,N5bis[pyridine-2-methylene]-thiocarbohydrazone Schiff-base Ligand. Spectrochim Acta 79:1050-1056.
- Ullah R, Ahmad I, Zheng Y (2016). Fourier Transform Infrared Spectroscopy of "Bisphenol A". Journal of Spectroscopy pp.1-5.

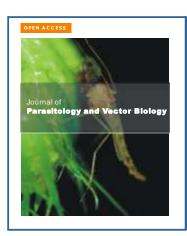
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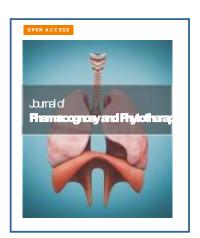


















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